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Research Article



METHOD DEVELOPMENT AND VALIDATION FOR ESTIMATION OF FAVIPIRAVIR BY USING RP-HPLC

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ABSTRACT:

A robust and precise analytical method was developed and validated through RP – HPLC for the estimation of Favipiravir by using a Discovery 250mm x 4.6 mm, 5m column with Acetonitrile: 0.01N Kh2po4 (70:30 v/v) as mobile phase. The method demonstrated excellent linearity in the concentration ranges of and 10–60 μ g/ml for Favipiravir with correlation coefficients (R²) of 0.999 for both drugs, confirming strong linearity. The regression equations obtained were and y = 63434x + 13743 for Favipiravir. The assay results indicated high accuracy with mean assay values of 99.46% for Favipiravir. Specificity studies confirmed the method's ability to distinctly quantify both analytes without interference. System suitability and method precision showed %RSD values within acceptable limits (\leq 2%), ensuring reproducibility. Accuracy studies yielded recoveries of 99.54% for Favipiravir. The sensitivity of the method was demonstrated by low limits of detection (LOD) and quantification (LOQ), observed at 0.03 μ g/ml and 0.10 μ g/ml for Favipiravir, respectively. Robustness studies further confirmed method reliability under small variations in method parameters

Key Words: Favipiravir, RP-HPLC, validation, Method Development.

Introduction:

Favipiravir (T-705, 6-fuoro-3-hydroxypyrazine-2carboxamide) is a new influenza medication that specifically inhibits the virus's RNA-dependent RNA polymerase.¹ Favipiravir is one of the prospective medications that might be employed in the treatment of COVID-19 infection.² Favipiravir was initially used against SARS-CoV-2 in Wuhan, the pandemic's epicenter. Then, when the epidemic expanded to Europe, this medicine got permission for emergency use in Italy and is now in use in other Countries.³ Favipiravir begins as a prodrug ribosylated and phosphorylated intracellularly to form the active favipiravir-RTP. 4,5 Favipiravir-RTP binds to and inhibits RNA dependent RNA polymerase (RdRp), effectively stopping viral transcription and replication.^{4,6}

Favipiravir is chemically known as 6-fluoro-3-hydroxypyrazine-2-carboxamide7 and The bioavailability of favipiravir is almost complete at 97.6%. It is converted inside cells into an active form known as favipiravir-ribofuranosyl-5'-triphosphate (favipiravir-RTP). ^{8,9} This active form is recognized by viral RNA-dependent RNA

polymerase (RdRp), an enzyme required for viral replication, and functions as a substrate, limiting the polymerase's activity. By disrupting the production of viral RNA, favipiravir prevents the virus from reproducing. ^{10, 11}

Figure 1: Structure of Favipiravir

High-performance liquid chromatography (HPLC) is critical in the analysis, method development, and validation of Favipiravir because it provides a dependable, selective, and sensitive platform for quantification in bulk drugs and pharmaceutical formulations. HPLC, particularly RP-HPLC, provides the effective separation of Favipiravir from its contaminants, excipients, and degradation

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products, making it critical for developing stability-indicating procedures. HPLC achieves precise peak resolution and repeatable retention characteristics by adjusting chromatographic settings such as mobile phase composition, pH, stationary phase, flow rate, and detection wavelength. During method validation, HPLC is essential for routine quality control, stability testing, impurity profiling, and confirming the safety and efficacy of Favipiravir drugs.

Extensive literature research has unearthed a multitude of recorded analytical procedures, including the discovery of more economically efficient ways¹²⁻²⁰. Hence, a reliable and cost-effective approach is suggested for assessing the stability of Favipiravir, and their medicinal dose form using RP-HPLC. must be validated and developed as per ICH guidelines

Materials and Methods:

Favipiravir (API), Favipiravir tablets (Fabiflu), Acetonitrile, Methanol, Ortho Phosphoric Acid, Distilled water. All of the solvents and chemicals were of HPLC quality and obtained from Rankem Chemicals Pvt Ltd.

Instrumentation:

The Method Development and Validation was performed by Waters HPLC Model 2695 equipped with PDA Detector and Empower 3 Software. Analytical weighing Balance, Ultrasonicator, pH Meter, Hot air oven.

Chromatographic Condition:

An Isocratic Elution carried out by using **Acetonitrile and 0.01N Kh2po4 30:70 v/v** as the Mobile Phase, Diluent used was Combination of Acetonitrile and Water in 1:1 ratio. **Discovery C18** (4.6 x 250mm, 5 μ m) column was used to determine the Method at a flow rate of 1ml/min, by maintaining the column Temperature at 30 OC. In addition, with an injection volume of 10 μ L and the wavelength detected at 320nm.

API Formulation

Preparation of Standard stock solutions and Working Solution: Accurately weighed 20mg of Favipiravir transferred to 50ml volumetric flask and 3/4 th of diluents was added to these flask and sonicated for 10 minutes. Flask were made up with diluents and labeled as Standard stock solution. (400μg/ml of Favipiravir). Moreover, from this 1ml from stock solution was pipetted out, taken into a 10ml volumetric flask, and made up with diluent to make 40μg/ml of Favipiravir.

Sample Formulation

Preparation of Sample stock solutions and Working Solution: 10 tablets were weighed and

the average weight of each tablet was calculated, then the weight equivalent to 1 tablet was transferred into a 100ml volumetric flask, 80ml of diluents was added and sonicated for 25 min, further the volume was made up with diluent and filtered by HPLC filters to make 2000µg/ml of Favipiravir solution. From this 0.2ml of filtered sample stock solution was transferred to 10ml volumetric flask and made up with diluent to make 40µg/ml of Favipiravir.

Method Validation

The established technique is validated in accordance with ICH criteria for the purpose of validating analytical methods. The validation metrics were: system appropriateness, accuracy, linearity, limit of detection (LOD), limit of quantification (LOQ), precision, robustness, specificity, and degradation studies.

System suitability parameters:

The chromatographic analysis was performed in accordance with the developed and optimized parameters after the working standard solution was introduced into the HPLC system six times. By calculating the % RSD of retention times, theoretical plates, and peak areas from six duplicate injections, the system suitability parameters were established.

Specificity: Checking of the interference in the optimized method. We should not find interfering peaks in blank and placebo at retention times of these drugs in this method. So this method was said to be specific.

Precision:

The procedure and system accuracy were demonstrated by creating a standard solution with a known concentration and providing six repeating injections to ensure the suggested approach's consistency. Intermediate precision was also attained by producing six working sample solutions and injecting them six times. The area was measured, and the mean, standard deviation, and % RSD were calculated. The results were favourable, falling below the 2% limit.

Linearity:

To test the drug's linearity, serial dilutions from 10 to 60 μ g/ml were prepared. A graph was used to demonstrate the link between peak area response and medication concentration. It was found to be linear at the indicated drug concentration. Dilution were as follows.

- 25 μg/mL: Take 0.25 mL of stock solution and dilute to 10 mL
- 50 μ g/mL: Take 0.5 mL of stock solution and dilute to 10 mL

- 75 µg/mL: Take 0.75 mL of stock solution and dilute to 10 mL
- 100 µg/mL: Take 1.0 mL of stock solution and dilute to 10 mL
- 125 μg/mL: Take 1.25 mL of stock solution and dilute to 10 mL
- 150 μ g/mL: Take 1.5 mL of stock solution and dilute to 10 mL

Accuracy:

Accuracy was performed in triplicate for various concentrations of Favipiravir equivalent to 50%, 100% and 150% of the standard amount were injected into the HPLC system per the test procedure. Dilution were as follows.

- 50 μg/mL: Take 0.1 mL of stock solution and dilute to 10 mL
- 100 μg/mL: Take 0.2 mL of stock solution and dilute to 10 mL
- 150 μ g/mL: Take 0.3 mL of stock solution and dilute to 10 mL

Sensitivity:

Limit of detection and Limit of Quantification

LOD and LOQ were calculated from the average slope and standard deviation from the calibration curve as per ICH guidelines.

Based on the response's standard deviation and calibration curve's slope, the LOD and LOQ can be estimated. The formulae given below can be used to calculate LOD and LOO:

$$LOD = 3.3\sigma/S$$

$$LOQ = 10\sigma/S$$

where S is calibration curve of the slope and σ is the response of the standard deviation.

Sensitivity Stock solution: Take 0.25 mL of stock solution and dilute to 10 mL

- LOD: From above take 0.3 ml solution and dilute to 10 mL
- LOQ: From above take 0.9 ml solution and dilute to 10 mL

Robustness: Small deliberate changes in method like Flow rate, mobile phase ratio, and temperature are made but there were no recognized change in the result and are within range as per ICH Guidelines.

Assay:

The assay and % purity were calculated for brand FabiFlu with label claim 200g. The observed value was compared with that of standard value without interference from the excipients used in the tablet dosage form

Degradation Studies

These investigations are carried out under various stress situations to describe the stability of the pure pharmacological material and are useful in establishing the best storage settings. These research cover base, peroxide, acid, neutral hydrolysis, photo, and heat degradation.

Oxidation:

After adding 1ml of a stock solution to 10ml of a 20% volume fraction of H2O2 and allowing it to sit in an oven at 60° C for 30 minutes, a chromatogram was produced by injecting a 40 μ g/ml solution at 10 μ l into HPLC

Acid Degradation Studies:

1 ml of hydrochloric acid was added to 10 ml of vf with 1 ml stock and refluxed for 30 minutes at 60 oC. A 40 μ g/ml solution was injected at 10 μ l into the system, resulting in the formation of a chromatogram.

Alkali Degradation Studies:

A mixture of 1 ml of stock and 1 ml of NaOH in 10 ml of vf was refluxed for 30 minutes at 60° C. A 40 μ g/ml solution was injected at 10 μ l into HPLC, resulting in the production of a chromatogram.

Dry Heat Degradation Studies:

The stock solution was allowed to undergo thermal deterioration in an oven set at 105 oC for 6 hours. Subsequently, a chromatogram was prepared by injecting a 40 µg/ml solution at 10 µl into HPLC.

Photo Stability studies:

The stock underwent degradation by exposure to UV radiation in the laboratory for a duration of 7 days. Upon injecting a 40 μ g/ml solution at a volume of 10 μ l into HPLC, a chromatogram was developed.

Neutral Degradation Studies:

After refluxing the stock for 6 hours at 60 degrees Celsius, a chromatogram was prepared by injecting a 40 µg/ml solution at 10 µl into HPLC.

RESULT AND DISCUSSION

Table 1 Optimized Condition

Parameter	Condition
Mobile phase	Acetonitrile: $0.01N \text{ KH}_2PO_4 (70:30 \text{ v/v})$
Flow rate	1 ml/min
Column	Discovery C18 (4.6 x 250mm, 5μm)
Detector wave length	320nm
Column temperature	30°C
Injection volume	10μL
Diluent	Water and Acetonitrile in the ratio 50:50

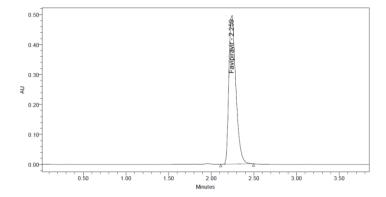


Figure 2: Optimized Chromatogram

System Suitability:

Table 2: System Suitability

S no			Favipiravir	
Injection	RT	area	Plate Count	Tailing
Injection-1	2.226	2563952	3289	1.42
Injection-2	2.229	2562960	3278	1.41
Injection-3	2.229	2560444	3250	1.41
Injection-4	2.232	2582241	3161	1.43
Injection-5	2.236	2565395	3213	1.43
Injection-6	2.238	2569317	3209	1.42
Mean		2567385		
Std ev		7847.8		
RSD		0.3		

Six injections of the standard Favipiravir solution were performed, and the chromatograms that corresponded to each injection were acquired. Observations showed that the percent RSD was less than 2%, the USP tailing was less than 2, and the theoretical plate count surpassed 2,000. Every condition for system appropriateness was satisfied and falls within permissible bounds.

Linearity:

Six concentrations ranging from 10 to 60 μ g/ml were prepared and linearity was estimated in a duplicate manner. The linearity equation for Favipiravir was y=63434x+13743. For the calibration curve over the concentration range, the data have shown a good correlation.

Table 3: Linearity Data

Concentration (ppm)	*Peak area
0	0
5	646906
10	1285168
15	1926182
20	2565364
25	3205994
30	3787705
<i>y</i> :	63434x+13743
R^2	0.999
Slope	63434
Intercept	13743
LOD	$0.03\mu g/ml$
LOQ	$0.10~\mu g/ml$

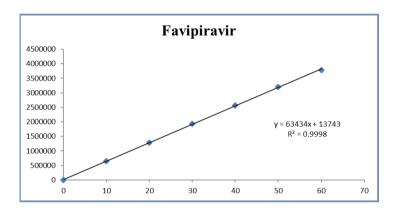


Figure 3: Calibration Curve of Favipiravir

Accuracy:

Three doses were given at each level, and the mean % recovery was calculated. Favipiravir's recovery rate was observed to be between 99% and 100.%, which is within the acceptable ranges

Table 4: Accuracy Data

% Level	Amount Spiked (µg/mL)	Amount recovered (µg/mL)	% Recovery	Avg %	Mean %Recovery
		19.93	99.67		
50%	20	19.89	99.47	99.62	
		19.94	99.71		
		39.94	99.85		00 = 40/
100%	40	39.90	99.76	99.87	99.54% %RSD: 0.33
		40.00	100.01		/0K3D: 0.33
		59.48	99.14		
150%	60	59.49	99.15	99.14	
		59.48	99.14		

Precision:

Table 5: Precision Data

S. No	Day 1	Day 2
Injection-1	2546513	2544421
Injection-2	2553840	2546924
Injection-3	2563781	2533857
Injection-4	2554472	2530194
Injection-5	2568937	2518236
Injection-6	2563648	2534963
Mean	2558532	2534766
S.D	8306.0	10354.2
%RSD	0.3	0.4

Robustness:

Table 6: Robustness data

condition	Optimized condition	Condition applied	area	RT	Plate Count	Tailing factor	RSD
Flow rate (-0.1ml/min)		0.9ml/min	2594426	2.363	3570	1.41	0.3
Mobile phase (-5%v/v)		35: 65	2600711	2.330	3468	1.44	0.3
Column temp (-3°c)	1ml/min	$27^{0}c$	2493810	2.335	3335	1.44	0.5
Optimized	30:70		2560334	2.259	3375	1.4	-
Flow rate (+0.1ml/min)	$30^{o}c$	1.1ml/min	2467774	2.088	3338	1.39	0.6
Mobile phase (+5%v/v)		25: 75	2535038	2.094	3207	1.42	0.5
Column temp $(+3^0c)$		$33^{0}c$	2608858	2.214	3392	1.44	0.3

Assay

Table 7: % Assay Purity Data

Formulation	Label claim(mg)	% Assay*
Fabiflu	Favipiravir 200 mg	99.46 %w/w

Degradation studies:

Table 8: Force Degradation Studies

S.No	% Degradation	Peak area	Stress Conditions	Peak Purity
1	6.24	2411441	Acid	Passes
2	3.88	2472137	Base	Passes
3	4.50	2456247	Oxidation	Passes
4	2.96	2495913	Thermal	Passes
5	1.21	2540792	Photolytic	Passes
6	0.85	2550111	Hydrolytic	Passes

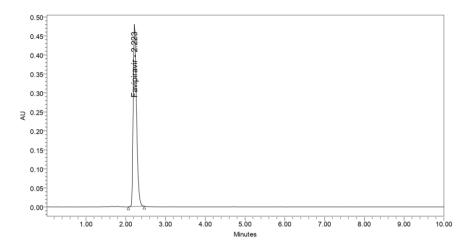


Figure 4: Acid Chromatogram

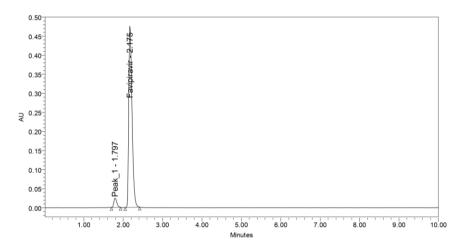


Figure 5: Base Chromatogram

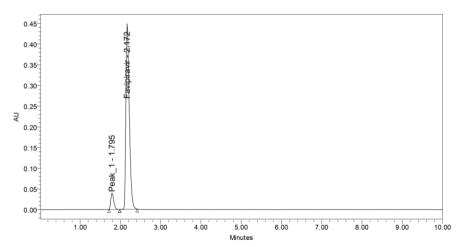


Figure 5: Oxidative Chromatogram

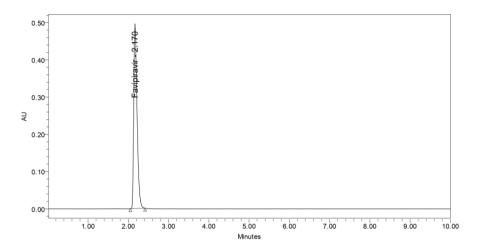


Figure 5: Thermal Chromatogram

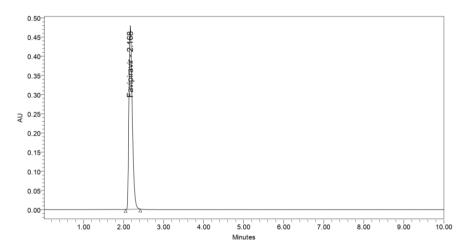


Figure 5: Photolytic Chromatogram

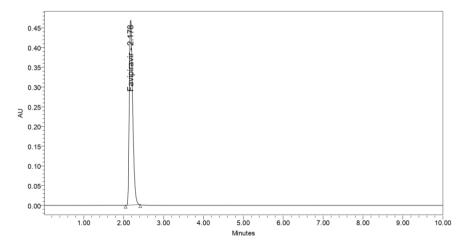


Figure 5: Water Chromatogram

CONCLUSION:

In summary, the Favipiravir HPLC analysis findings demonstrate that this method can accurately ascertain the drug's concentration and purity. The technique is perfect for routine quality control and pharmacokinetic research because of its strong repeatability, clear peak resolutions, and consistent retention times. HPLC is a crucial method for evaluating the analytical qualities of favipiravir in order to guarantee its effectiveness and safety for therapeutic usage.

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